

1 **Flower visitation through the lens: Exploring the foraging behaviour of *Bombus***  
2 ***terrestris* with a computer vision-based application**

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18 interactions, flower visitation, deep learning, computer vision, YOLO

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20

## 21 Abstract

22 To understand the processes behind pollinator declines, and thus to maintain pollination  
23 efficiency, we also have to understand fundamental drivers influencing pollinator behaviour. In  
24 this study, we aim to explore the foraging behaviour of wild bumblebees, recognizing its  
25 importance from economic and conservation perspectives. We recorded *Bombus terrestris* on  
26 *Lotus creticus*, *Persicaria capitata*, and *Trifolium pratense* patches in five-minute-long slots in  
27 urban areas of Terceira (Azores, Portugal). For the automated bumblebee detection, we created  
28 computer vision models based on a deep learning algorithm, with custom datasets. We achieved  
29 high F1 scores of 0.88 for *Lotus* and *Persicaria*, and 0.95 for *Trifolium*, indicating accurate  
30 bumblebee detection. We found that flower cover per cent, but not plant species, influenced the  
31 attractiveness of flower patches, with a significant positive effect. There were no differences  
32 between plant species in the attractiveness of the flower heads. The handling time was longer  
33 on the large-headed *Trifolium* than those on the smaller-headed *Lotus* and *Persicaria*. However,  
34 our result did not indicate significant differences in the time bumblebees spent on flowers  
35 among the three plant species. Here, we also justify computer vision-based analysis as a reliable  
36 tool for studying pollinator behavioural ecology.

37

## 38 1. Introduction

39 In the Anthropocene, the populations of native pollinators are declining worldwide (Nath et al.,  
40 2023; Potts et al., 2010), posing a threat to pollinator-dependent crops and wildflowers  
41 (Biesmeijer, 2006; Vanbergen & Initiative, 2013). This, in turn, hampers ecosystem functioning  
42 of natural and agroecosystems and jeopardises the delivery of ecosystem services vital for  
43 humans. While key pressure on pollinators influencing this decline, such as climate change  
44 (Kerr et al., 2015; Martinet et al., 2020), land-use change, habitat loss or fragmentation  
45 (Vanbergen, 2014), and pesticide inputs (Godfray et al., 2014; Stanley et al., 2015), are well  
46 studied (Dicks et al., 2021), their effects on the behaviour of pollinators remain unexplored.  
47 Indeed, the effective composition of pollinator communities not only changes with the  
48 disappearance of species but also with the subtle behavioural changes, that often go undetected,  
49 as the remaining species adapt to new conditions (Kaiser-Bunbury et al., 2010; Schweiger et  
50 al., 2010). Since these changes can negatively impact behaviour-mediated ecosystem functions  
51 such as pollination (Lippert et al., 2021), to slow down or stop negative population trends and  
52 promote recovery we need to understand previously unexplored and intricate details of plant-  
53 pollinator interactions, including those related to pollination behaviour (Burkle & Alarcón,  
54 2011; Byers, 2017).

55 In addition to the undeniable importance of domesticated honeybees (*Apis* sp.) in farming,  
56 native and domesticated bumblebees (*Bombus* sp.) play a significant role not only in pollinating  
57 wildflowers in natural ecosystems but also in agricultural crop production by maintaining high  
58 yields (Rao & Stephen, 2009; Velthuis & Doorn, 2006). Indeed, among wild bees, bumblebees  
59 are the highest contributors to crop pollination (Kleijn et al., 2015; Ollerton et al., 2011). Yet,  
60 they are particularly exposed to the effects responsible for global pollinator declines (Goulson  
61 et al., 2008; Soroye et al., 2020; Williams & Osborne, 2009), and their populations have also  
62 been reported to show steep declining trends which were predicted to continue and even  
63 accelerate for many species (Ghisbain et al., 2023; Nieto, 2014). This decline is exacerbated by

64 changes in bumblebee behaviour due to various anthropogenic and biotic impacts. For instance  
65 widely used agrochemicals and viral or parasite infections can cause non-lethal changes (Varga-  
66 Szilay & Tóth, 2022), such as reduced homing ability, colony growth (Stanley et al., 2016), and  
67 food intake capacity (Feltham et al., 2014) of the individuals, through which the existence of  
68 entire colonies can be jeopardized.

69 One of the most important behaviours likely to be prone to changes is foraging, which is vital  
70 in driving the fitness of bumblebee populations, and thus long-term pollination efficiency. Since  
71 it is of high economic and conservation importance, numerous studies scrutinised this behaviour  
72 and found that the foraging success and homing ability of bumblebees can be influenced by a  
73 multitude of factors. Large-scale abiotic factors such as temperature, humidity, or pesticide  
74 exposure could affect flower handling and spatial foraging behaviour (Gill et al., 2012;  
75 Samuelson et al., 2016). Small-scale external factors include flower morphology and colour,  
76 the characteristics of flowering patches, parasites (Gillespie & Adler, 2013), the cost of flight  
77 between patches (Goulson, 2000), forager density, and the spatial distribution of flowers  
78 (Geslin et al., 2014). Internal factors, such as learning abilities (Evans et al., 2017) and physical  
79 conditions of foragers (Gegear et al., 2006), also play a role. Despite these influences,  
80 bumblebees show high behavioural plasticity and forage efficiently under a wide range of  
81 environmental conditions (Goulson, 2010; Jha & Kremen, 2013; Zimmerman, 1981) making  
82 them less constrained in their foraging behaviour than other insects (Goulson, 2010). To  
83 appropriately assess anthropogenic impacts, though, a mechanistic understanding of  
84 fundamental foraging behavioural patterns, such as how resource handling times and visiting  
85 frequencies depend on plant species or flower density-dependent carrying capacity of patches  
86 is needed (Jha & Kremen, 2013). Yet, key information on these important aspects of bumblebee  
87 ecology is scarce, most likely because of the laborious means of data collection. In fact, whilst  
88 there is a large set of tools for studying pollinator community ecology, high-throughput methods

89 for effectively monitoring behavioural changes of wild bumblebee populations are still missing,  
90 which substantially hamper study efforts.

91 Traditionally used observational methods for recording pollinators' behaviour and activity, such  
92 as transects, mark-recapture, and timed count-based observations, are not only time-consuming  
93 and difficult to standardise (Darras et al., 2019) because they highly depend on the skills of the  
94 person conducting the field observations, but they also can disturb the insects' natural behaviour  
95 and thus skew the result (Besson et al., 2022; Tuia et al., 2022). Although standardised  
96 behavioural observations under laboratory conditions, at least partially, address these issues,  
97 they are challenging to adapt to wild conditions, especially because these studies are often  
98 conducted on commercially produced bumblebees rather than wild populations (Treanore et al.,  
99 2021).

100 However, modern methods, such as using video recording to observe the foraging behaviour of  
101 insect floral visitors around flower sources present an opportunity for increasing efficiency,  
102 decreasing disturbance, and saving human labour. When combined with state-of-the-art  
103 technologies, including computer vision and deep learning techniques, video recordings can  
104 provide novel solutions to species identification (Bjerge et al., 2023; Spiesman et al., 2021) and  
105 monitoring communities (Besson et al., 2022) or insect pests (Prete et al., 2021). Indeed, these  
106 methods are increasingly used in pollination ecology to detect flower-visiting insects (e.g.  
107 (Ratnayake et al., 2022; Sittinger et al., 2024)) and recent trends suggest that the future of  
108 pollinator research is going to be shaped by the use of artificial intelligence (AI)-based tools  
109 (Barlow & O'Neill, 2020; Høye et al., 2021; van Klink et al., 2022). Despite their numerous  
110 advantages, few studies use these methods to monitor pollinators in real-time (but see (Bjerge  
111 et al., 2021; Ngo et al., 2021)) and even less to examine unmarked insects behaviour outdoors,  
112 under natural or near-natural conditions (but see (Bjerge et al., 2022; Ratnayake et al., 2021)).  
113 One of the reasons may be that, whilst the application of AI tools rarely needs any specialised

114 technological hardware, at their current development stage, these methods often require specific  
115 computational skills, which extend beyond the expertise of most ecologists. The intensive use  
116 of a non-ecology-related discipline thus highlights the need for enhancing collaboration  
117 between ecologists and computer scientists (Carey et al., 2019).

118 Additionally, the effectiveness of computer vision and object detection in field settings, and  
119 thus the success of data extraction, depends on several environmental factors such as light  
120 conditions, wind strength, and precipitation. To overcome these challenges, some studies used  
121 artificial platforms (Sittinger et al., 2024) or fixed flower heads to sturdy surfaces (Steen, 2017).  
122 However, no studies have yet been conducted to assess pollinator behavioural patterns using  
123 computer vision in completely uncontrolled field settings.

124 In this research, we used computer-vision-based methods to study wild populations of buff-  
125 tailed bumblebees (*Bombus terrestris* (Linnaeus, 1758), Hymenoptera, Apidae) foraging on  
126 three wild-growing, insect-pollinated plants: Cretan bird's-foot trefoil (*Lotus creticus* Linnaeus,  
127 1753, Fabales, Fabaceae), pink-headed knotweed (*Persicaria capitata* (Buch.-Ham. ex D.Don)  
128 H.Gross, Caryophyllales, Polygonaceae), and red clover (*Trifolium pratense* L., Fabales,  
129 Fabaceae) in urban areas. This work aimed to explore and understand the foraging behaviour  
130 of bumblebees under field conditions and to describe the differences between their behavioural  
131 patterns on the different flower species and with varying flower densities. At the same time, our  
132 alternative objective was to test whether video-based recording methods combined with  
133 computer vision-based analysis are suitable for exploring bumblebee behaviour under field  
134 conditions.

135 We aimed to answer the following questions:

- 136 • Question 1: How many bumblebees a flower patch can support within a specified time  
137 unit? Is there a difference in the visitation of flower patches per time unit?
- 138 • Question 2: How many bumblebees can occupy a flower patch simultaneously?

139 • Question 3: What proportion of their total time (*'bumblebee-time'*) bumblebees spend  
140 on flowers (handling time) compared to time spent on non-flowery areas (travelling  
141 time)?

142 Our hypothesis was that the assumed optimal foraging behaviour of the nectar- and pollen-  
143 gathering bumblebees show significant differences in the time spent on flowers among the three  
144 plant species and that it is adjusted to the characteristics of the resources (three types and density  
145 of flower patches and flower head size of plants).

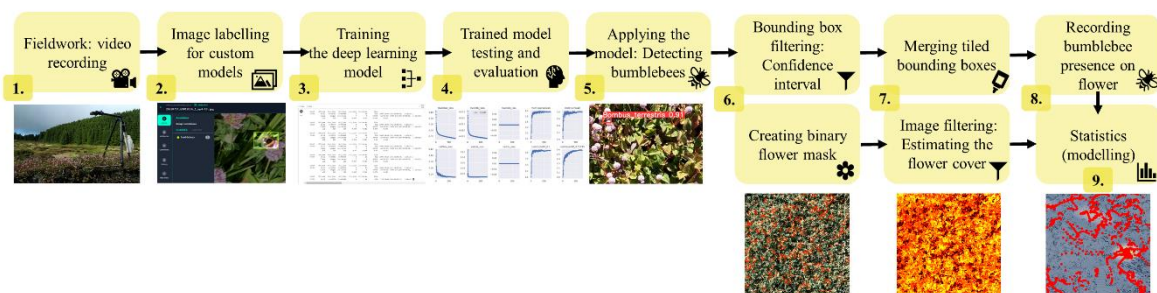
146

## 147 2. Material and methods

### 148 1.1. Study sites

149 The study was conducted on Terceira Island (Azores, Portugal) between May and September  
150 2022. The sampling sites were located in urban areas at 38°48'06.0" N, 27°15'17.8" W,  
151 38°44'14.7" N, 27°16'07.8" W, and 38°47'38.7" N, 27°15'24.0" W (**Figure S1**). We recorded  
152 the bumblebees (*Bombus terrestris*) on Cretan bird's-foot trefoil (*Lotus creticus*; indeterminate  
153 biogeographic origin), pink-headed knotweed (*Persicaria capitata*; introduced invasive), and  
154 red clover (*Trifolium pratense*; introduced naturalised) patches.

155



156

157 **Figure 1:** Flowchart of the data collection, preparation and analysis to assess bumblebee  
158 behavioural differences of three plant species.

159

## 160 1.2. Data collection

161 Videos were recorded with GoPro Hero9 action cameras in approximately five-minute-long  
162 slots in 5K resolution (5120 by 2880) at 30 frames per second speed on 60x60 cm square  
163 quadrats (**Figure 1**). We excluded videos without bumblebees from the analysis (nine videos  
164 from *Lotus* and two from *Persicaria*), which resulted in 15, 18, and 15 videos from *Lotus*,  
165 *Trifolium*, and *Persicaria*, respectively.

166 Although we attempted to take the videos at the same height from the ground, this was not  
167 always possible due to the uneven surface. Therefore, to allow size and area estimations, each  
168 setup was calibrated with a millimetre-precision scale and the real-life length of one pixel was  
169 calculated. Metadata showing recording location, date and time was linked to each video and  
170 used in the analysis. Temperature (C°) and humidity (%) were measured, and wind strength  
171 (Beaufort Wind Scale) and cloudiness (direct sunshine, overcasted, or cloudy) were estimated  
172 on-site.

## 173 1.3. Data processing

174 Videos were split into frames and frame-level information was further used for training and  
175 analysis. For labelling the training set for the deep learning algorithm, three-second segments  
176 (90 frames) were cut from the beginning of each video or from the frame where the first  
177 bumblebee(s) appeared in the video. The images were manually annotated by drawing bounding  
178 boxes of one label class ('bumblebees') around the bumblebees with the help of Roboflow  
179 annotation tool (Dwyer et al., 2024). The datasets (*Lotus*: 4308 images (Varga-Szilay, 2023b),  
180 *Persicaria*: 1908 images (Varga-Szilay, 2023a), *Trifolium*: 2099 images (Varga-Szilay, 2023c))  
181 comprising annotated bumblebees, were split into training, cross-validation, and test sets (as  
182 70, 20, and 10% proportions, respectively, **Table 1**). A proportion of 5% of the original image  
183 number was also added as false positive images for each type of image set (**Table 1**).

184

185 **Table 1:** The dataset of images used to train the object detection model (YOLOv5). The table  
186 shows the number of 'bumblebee'-labelled images, as well as the false positive images (e.g. with  
187 background or shadows) without label.

| Type of set  | Type of images | Flower species |                  |                   |
|--------------|----------------|----------------|------------------|-------------------|
|              |                | <i>Lotus</i>   | <i>Trifolium</i> | <i>Persicaria</i> |
| Test         | Labeled        | 387            | 254              | 202               |
|              | False positive | 19             | 12               | 10                |
| Train        | Labeled        | 2889           | 68               | 1247              |
|              | False positive | 144            | 1354             | 62                |
| Validation   | Labeled        | 828            | 391              | 369               |
|              | False positive | 41             | 20               | 18                |
| <b>Total</b> |                | <b>4308</b>    | <b>2099</b>      | <b>1908</b>       |

188  
189 For training, to keep the resolution high yet allow tiling with 640-pixel (px) segments, the  
190 images were expanded from 5120x2880 to 5120x3200 and then they were cropped onto 640 x  
191 640 px tiles (pre-processing). For the automated detection of bumblebees in the videos we  
192 created deep learning-based computer vision models for each plant species separately, using  
193 YOLOv5 (You Only Look Once (Jocher, 2020)) with custom datasets for each plant species.  
194 For the training process we used either Google Colab (Tesla 4T 15102MiB GPU) or a desktop  
195 PC (11<sup>th</sup> Gen Intel(R) Core(TM) i9/11900KF @ 3.50GHz, 64 GB RAM, 8 Cores, Win11,  
196 NVIDIA GeForce RTX 3060, 12288 MB) with a PyTorch (Paszke et al., 2019) environment  
197 with 0.01 learning rate (LR). All models were trained to 300 epochs with 64 batch size and  
198 default hyperparameters. See the specific evaluation metrics of YOLOv5 models in **Figure S2**.  
199 Further analysis was performed with a data set (bounding box set) filtered twice to minimize  
200 false bumblebee detections. For bounding box filtering, a confidence level of 0.7 was used for  
201 the object detection results for *Trifolium*, and 0.8 for *Lotus* and *Persicaria*. To avoid multiple  
202 detections of one bumblebee through the tiling process after the tiles were merged we calculated  
203 the Euclidean distance between each bounding box and merged the boxes if the distance  
204 between them was less than or equal to the size of half of an average bumblebee (15 mm  
205 calculated from the calibrated pixel sizes) (post-processing). To test the post-processing

206 accuracy of the model on filtered bounding boxes, we randomly selected five frames from five  
207 videos for each flower species and compared the manual detections with those predicted by the  
208 model (**Table S1**).

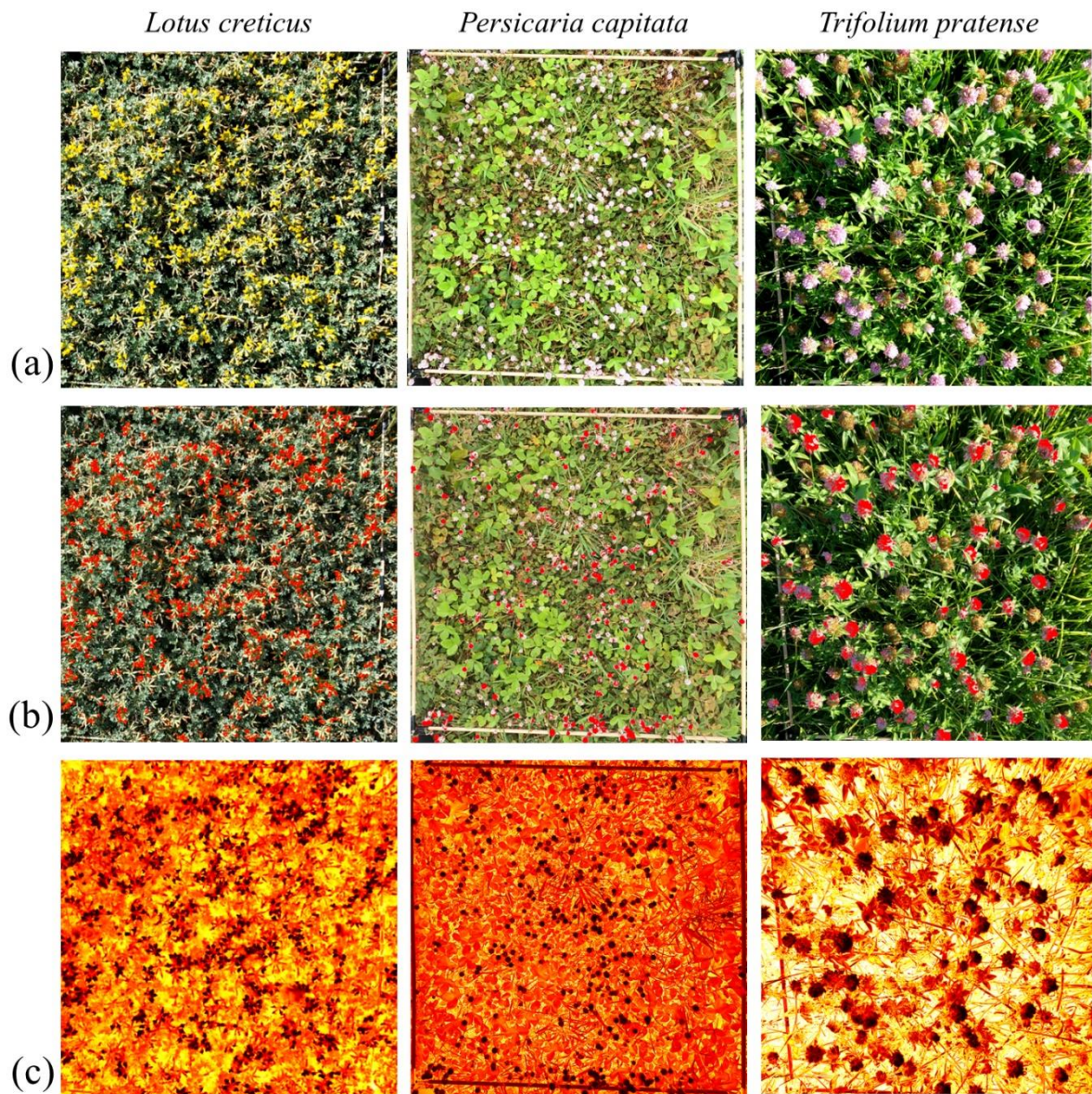
209 To crop the quadrat from the video, the upper left corner of the physically placed square quadrat  
210 was digitally identified, and the  $x1$  and  $y1$  coordinates of this corner were recorded. Then, the  
211 remaining coordinates ( $x2$ ,  $y2$ ) of the 60 x 60 cm square were computed knowing the one  
212 pixel/mm value. The quadrats were then extracted from each video frame based on these  
213 calculated coordinates. Bumblebee detections were only kept if the centroids of the bounding  
214 boxes were within the quadrat.

215 For flower detection, we manually determined the flower species-specific upper and lower Hue,  
216 Saturation and Brightness (HSB) colour thresholds from the unedited images (**Figure 2a**) with  
217 the ImageJ software (Rasband, 2018)) (*Lotus*: 11, 42, 120, 255, 160, 255; *Persicaria*: 140, 255,  
218 0, 140, 40, 255; *Trifolium*: 85, 255, 0, 247, 186, 255) and used these in a colour filtering process.  
219 The ImageJ-compatible HSB ranges were later converted to Python-compatible Hue, Saturation  
220 and Brightness (HVS) colour thresholds. To calculate an average flower colour within this  
221 range, the ‘optimal flower colour’ (OFC), ten random frames were selected from each video  
222 and a colour mask for the HVS colour thresholds was applied to each of these frames (**Figure**  
223 **2b**). Then the median of all of these colour masks was determined for each flower species. To  
224 estimate the per cent cover of flowers (henceforth flower cover) in the quadrat, we created a  
225 binary mask based on the HSV range, counted the number of masked pixels and then the  
226 proportion of flower area in a quadrat was calculated by dividing this number by the total  
227 number of pixels in the quadrat. The value was calculated from one random frame for each  
228 video separately.

229 To create the heatmap, we utilized average similarity from the OFC of 3000 frames per video.

230 To get this colour similarity from the predefined OFC we calculated the distance between each

231 pixel in an HSV image (= one frame) and the OFC by extracting the Hue, Saturation, and Value  
232 components, then computing the Euclidean distance, taking into account the circular nature of  
233 the Hue component (**Figure 2c**). The areas of bounding boxes marking bumblebees were  
234 excluded from the average calculation.



235  
236 **Figure 2:** Example of (a) original images, (b) images filtering with binary mask, and (c)  
237 heatmap-based images indicating the Euclidean distance of the actual colour from the ‘optimal  
238 flower colour’ (brighter red indicates greater distance).

239

#### 240 1.4. Statistical analysis

241 To visualise the frequency of pixel colour distances from the optimal flower colour on a  
242 histogram for the flower species, we took a random sample (100,000 points or as many as were  
243 available) from areas on the heatmap where bounding boxes were and from the whole heatmap  
244 (full quadrat).

245 To determine how many bumblebees were attracted to each flower patch (for **Question 1**), we  
246 calculated the sum number of bumblebees per video divided by the total number of frames in  
247 the video and used this measure as a proxy for the attractiveness of a flower patch. When this  
248 number was standardized by the flower cover too (assuming that bees are more likely to handle  
249 flowers when flower sources are more abundant) we got a proxy for the carrying capacity of  
250 each plant species (for **Question 2**).

251 To determine whether bumblebees were on flowers, we used the flower colour thresholds to  
252 mask the bounding box area enclosing a bumblebee from a frame in which the animal was not  
253 present (to see the flower colours rather than those of the bumblebee's) and calculated the  
254 proportion of these masked pixels relative to the total pixels of the bounding box area. If the  
255 masked pixels (indicating flower colours) reached 20%, we considered the bumblebee to be on  
256 a flower at the moment of the detection. Thus, we refer to 'handling time' or 'on-flower time'  
257 when the detected bumblebee was on a flower, while bumblebee detections, where the insect  
258 was unlikely on a flower, are termed 'travelling time' or 'off-flower time'. To estimate a proxy  
259 for handling time (for **Question 3**), we calculated the proportion of 'bumblebee-on-flower-  
260 time' (summarising all time units (frames) that all detected individuals cumulatively spent on  
261 flowers) to all 'bumblebee-time' (that summarised all time units that all individuals,  
262 cumulatively, spent within the quadrat).

263 We used simple linear regression analysis and linear mixed models (wind strength and  
264 temperature as random effects) to evaluate the impact of plant species and flower cover on  
265 bumblebee behaviour. Mixed models were kept when they demonstrated superior performance

266 compared to the simple linear models, in terms of the Akaike Information Criterion (AIC), and  
267 the fit was not singular, otherwise, the simple linear regression models were used. All models  
268 were tested for homoscedasticity and the normality of residuals using the Shapiro-Wilk and the  
269 Goldfeld-Quandt tests, respectively. When assumptions of normality and homoscedasticity  
270 were not met, we either square-root or cubic-root transformed the response variables (**Table 2**).  
271 Heteroscedasticity was accounted for by applying heteroscedasticity-robust standard errors  
272 with the help of the ‘sandwich’ R package.

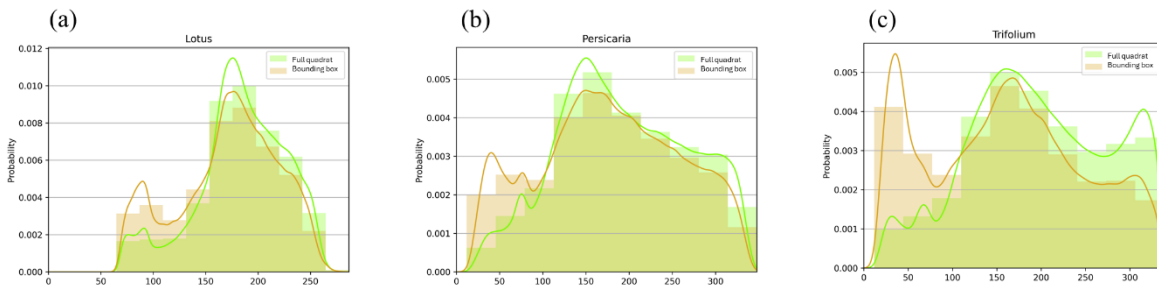
273 For data preparation and visualization, we used Python-3.8.16 (Python Software Foundation,  
274 2019) environment and torch-1.13.1, with the help of ‘NumPy’ version 1.23.5 (C. R. Harris et  
275 al., 2020), ‘Pandas’ version 1.5.3 (The pandas development team, 2022), ‘cv2’ version 4.7.0  
276 (Bradski, 2000), ‘ffmpegcv’ (FFmpeg Developers, 2016), ‘matplotlib’ version 3.7.1 (Hunter,  
277 2007), and ‘scipy’ version 1.10.0 (Virtanen et al., 2020) libraries. For data preparing, modelling  
278 and the visualisation of model results, we used the ‘dplyr’ (Wickham et al., 2023), ‘readr’  
279 (Wickham et al., 2024), ‘purrr’ (Wickham & Henry, 2023), ‘lmtest’ (Zeileis & Hothorn, 2002),  
280 ‘sandwich’ (Zeileis et al., 2020), and ‘ggplot2’ (Wickham, 2016) packages in an R environment  
281 (R version 4.4.0 (R Core Team, 2021)).

### 282 3. Results

283 We recorded 134,865 frames in 15 videos on *Lotus*, 154,151 frames in 18 videos on *Trifolium*,  
284 and 129,145 frames in 15 videos on *Persicaria*. The average percentage of flower cover was  
285 the lowest on *Persicaria* (2.86%,  $SD \pm 0.85$ ), while it was 4.48% ( $SD \pm 2.23$ ) and 5.29% ( $SD$   
286  $\pm 1.12$ ) on *Lotus* and *Trifolium*, respectively.

287 Higher F1 scores for *Lotus* and *Trifolium* indicated reliable overall performance of the model  
288 in accurately identifying and distinguishing between bumblebees compared to the detection on  
289 *Persicaria* (**Figure S3**). After the post-processing, the reliability of the F1 score further  
290 increased to 0.88 for the *Lotus* and *Persicaria* and 0.95 for the *Trifolium* (**Table S1**). Overall,

291 the model demonstrated high accuracy in finding the bumblebees and a low rate of false  
 292 detections. We recorded bumblebees within the quadrats on 177,271 occasions across 418,161  
 293 frames. The maximum number of bumblebee individuals at the same time on *Lotus* was 6 (mean  
 294 = 1.24, SD  $\pm$  0.26), on *Persicaria* it was 5 (mean = 1.16, SD  $\pm$  0.15), and on *Trifolium* it was 4  
 295 (mean = 1.07, SD  $\pm$  0.08). There was a difference in the handling time between different plants,  
 296 with the longest recorded being on *Trifolium* (**Figure 3**). In addition, bumblebees spent more  
 297 time off-flower on *Lotus* and *Persicaria* patches than on *Trifolium*.  
 298



299  
 300 **Figure 3:** The histograms show the distribution of Euclidean distances of the colours of sample  
 301 pixels from the predefined optimal flower colour (OFC = 0.0 on the x-axis) on *Lotus* (a),  
 302 *Persicaria* (b), and *Trifolium* (c) patches. Green coloured bars indicate pixels randomly selected  
 303 from the full quadrat, whilst the distribution of the colour deviation from OFC under bounding  
 304 boxes enclosing detected bumblebees is indicated in orange. The colour coding of the smoothed  
 305 density curves follows that of the bars.  
 306  
 307 **Table 2.:** Summary statistics of the linear regression and linear mixed models.

| <b>Question 1</b>   |              |           |                |                |
|---|--------------|-----------|----------------|----------------|
| Model: average number of bumblebees per time unit ~ flower species * flower cover |              |           |                |                |
|   | <b>slope</b> | <b>SD</b> | <b>t value</b> | <b>p-value</b> |
| (Intercept)   | 0.053        | 0.164     | 0.322          | 0.749          |
| <i>Persicaria</i>   | 0.205        | 0.304     | 0.675          | 0.504          |
| <i>Trifolium</i>  | -0.054       | 0.361     | -0.149         | 0.882          |
| Flower cover (%)  | 0.110        | 0.033     | 3.330          | <b>0.002**</b> |
| <i>Persicaria</i> x Flower cover (%)  | -0.069       | 0.092     | -0.744         | 0.461          |

|                                     |        |       |        |       |
|-------------------------------------|--------|-------|--------|-------|
| <i>Trifolium</i> x Flower cover (%) | -0.065 | 0.068 | -0.950 | 0.347 |
|-------------------------------------|--------|-------|--------|-------|

### Question 2

Model: (average number of bumblebees visiting flower patches at the same time)<sup>-3</sup> ~ flower species

|                   | slope   | SD     | t value | p-value   |
|-------------------|---------|--------|---------|-----------|
| (Intercept)       | 75.445  | 14.115 | 5.345   | <0.001*** |
| <i>Persicaria</i> | -56.155 | 19.963 | -2.813  | 0.007     |
| <i>Trifolium</i>  | 57.595  | 19.112 | 3.013   | 0.004     |

Model: (average number of bumblebees visiting flower patches at the same time)<sup>-3</sup> ~ flower species + (1 | wind strength) + (1 | temperature)

|                   | slope   | SD     | df    | t value | p-value |
|-------------------|---------|--------|-------|---------|---------|
| (Intercept)       | 1.359   | 53.919 | 4.432 | 0.025   | 0.981   |
| <i>Persicaria</i> | 58.949  | 70.381 | 4.834 | 0.838   | 0.442   |
| <i>Trifolium</i>  | 151.297 | 58.661 | 4.227 | 2.579   | 0.059   |

### Question 3

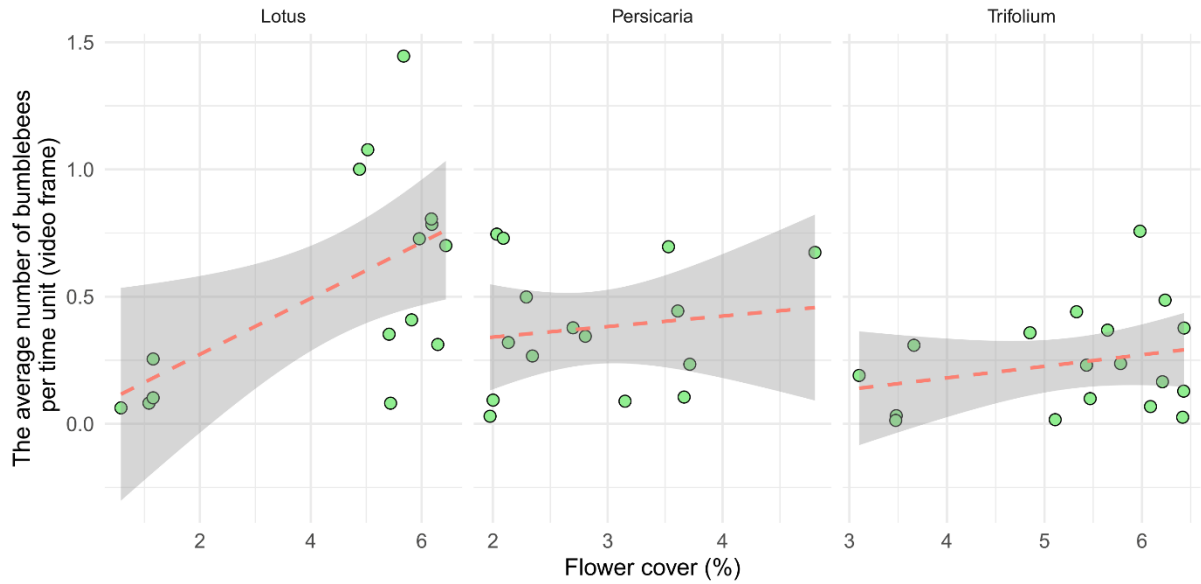
Model: percentage of 'bumblebee-time' spent on flower ~ flower species \* flower cover

|                                      | slope   | SD     | t value | p-value       |
|--------------------------------------|---------|--------|---------|---------------|
| (Intercept)                          | 30.861  | 18.875 | 1.635   | 0.110         |
| <i>Persicaria</i>                    | -32.511 | 20.561 | -1.581  | 0.121         |
| <i>Trifolium</i>                     | 19.119  | 35.137 | 0.544   | 0.589         |
| Flower cover (%)                     | 0.313   | 3.264  | 0.096   | 0.924         |
| <i>Persicaria</i> x Flower cover (%) | 8.385   | 4.104  | 2.043   | <b>0.047*</b> |
| <i>Trifolium</i> x Flower cover (%)  | -3.084  | 6.383  | -0.483  | 0.631         |

Indication of significance: 0 '\*\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

308

309 There were more bumblebees in *Lotus* than in the other two plant species within an average  
 310 time unit. The attractiveness of flowering patches was significantly influenced by flower cover  
 311 ( $p < 0.01$ , **Table 2**) but the plant species did not show significant effects (**Figure 4**). The curve  
 312 was steep in the case of *Lotus*, which may have been caused by the few low values of our  
 313 measure at extremely low cover values. However, the steep relationship between the cover and  
 314 bumblebee visitation on *Lotus* remained even after those outliers were removed.



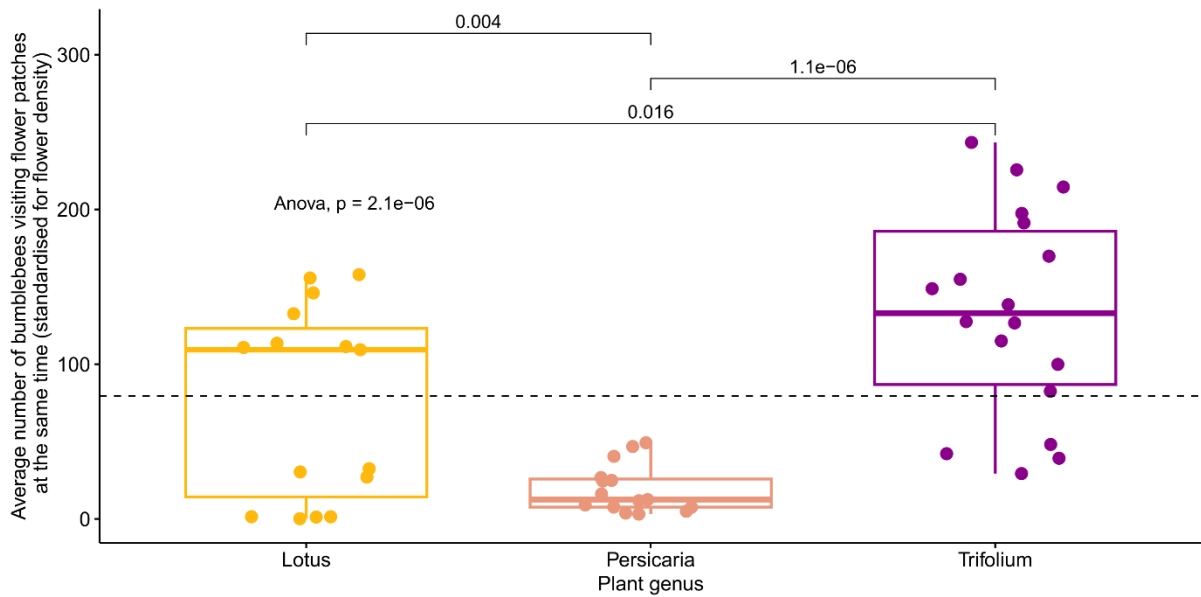
315

316 **Figure 4:** Visitation of flower patches per unit time (video frame) as a function of flower cover  
317 separated by plant species. Each point represents one video (n = 15, 15 and 18, for *Lotus*,  
318 *Persicaria*, and *Trifolium*, respectively).

319

320 When the plants' carrying capacity was investigated, the simple linear regression model showed  
321 a significant difference ( $p < 0.001$ ) between the flower species in the average number of  
322 bumblebees visiting flower patches at the same time (standardised for flower cover) (**Figure**  
323 **5**). However, when we controlled for wind strength and temperature in a random model no  
324 significant differences were found between the plant species (**Table 2, Figure S4**).

325



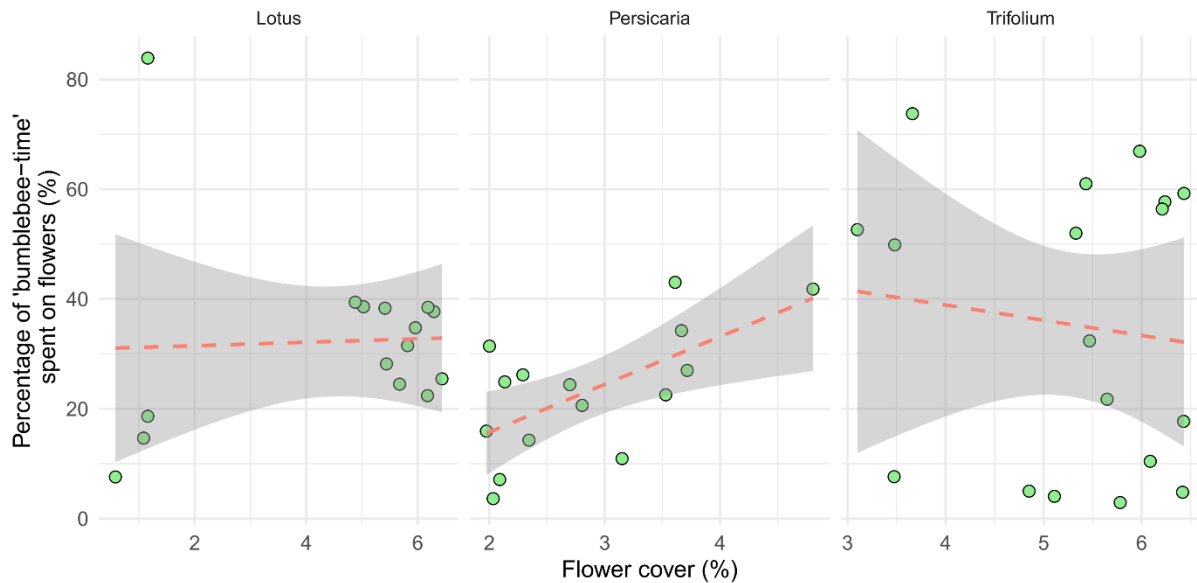
326

327 **Figure 5:** Average number of bumblebees visiting flower patches at the same time (standardised  
328 for flower cover) separated by plant species. The global p-value for the ANOVA test is shown  
329 in the figure, as are the pairwise comparisons (t-tests) of the averages between plant species.  
330 The dashed line shows the mean of the y-axis. Each point represents one video (n = 15, 15, and  
331 18, for *Lotus*, *Persicaria*, and *Trifolium*, respectively).

332

333 The model indicated a significant positive correlation between ‘*bumblebee-time*’ (individuals x  
334 time spent on flowers) spent on flowers and flower cover but only on *Persicaria* patches (p =  
335 0.047, **Table 2**). There was an indication of a slight but not significant negative interaction  
336 between ‘*bumblebee-time*’ spent on *Trifolium* flowers and flower cover (**Figure 6**).

337



338

339 **Figure 6:** The time bumblebees spent on flowers as a per cent of all time spent in the quadrat  
340 ('bumblebee-time'). Each point represents one video (n = 15, 15, and 18, for *Lotus*, *Persicaria*,  
341 and *Trifolium*, respectively).

342

#### 343 4. Discussion

344 In this study, we presented a computer-vision-based method to identify differences in the  
345 foraging behaviour of wild populations of bumblebees on three insect-pollinated plants among  
346 flower species and varying flower cover. We found only slight differences in the foraging  
347 behaviour of bumblebees among plant species but detected an indication of the importance of  
348 flower cover. This was likely because bumblebees adapted to the characteristics of the flower  
349 resources, including flower head sizes, as well as the different flower cover, which influenced  
350 their handling time and travelling behaviour.

##### 351 4.1. The attractiveness of the flower patch

352 We found that the number of bumblebees visiting a patch depended solely on the flower cover  
353 and not on the plant species. This aligns with the findings of Vaca-Urbe et al. (2021) (Vaca-  
354 Urbe et al., 2021) who reported a positive relationship between the blooming cover and the  
355 abundance of insect visitors. In our study, the observed dependency on flower cover was

356 particularly evident with *Lotus*, where patches with low flower cover had fewer bumblebees,  
357 while higher flower cover attracted a higher number of individuals, and this relationship  
358 between the flower cover and the number of bumblebees remained even after excluding outliers  
359 with extreme low cover values. One of the reasons for this is likely that in denser flower patches,  
360 bumblebees can spend more time handling flowers, minimizing the energy costs that would be  
361 spent searching for a new patch and maximizing resource uptake. However, the exact  
362 relationship between increasing cover and bumblebee visitation and its environmental drivers  
363 is challenging to determine, as we did not sample flower patches with intermediate flower  
364 cover.

365 Yet, other bumblebee species showed differences in their foraging behaviour, including their  
366 patch choice, based on flower cover or flower complexity (Stout et al., 1998).

#### 367 4.2. The carrying capacity of the flower species

368 The different results from the simple linear and the linear mixed model indicate no clear  
369 evidence of whether there is a difference in how many bumblebees can simultaneously occupy  
370 each flower species. Indeed, the variability of environmental parameters, such as temperature  
371 and humidity, both of which could affect the nectar and pollen production of the plants (C.  
372 Harris & Ratnieks, 2024), and the limited sample size may have masked differences among  
373 flower species. In addition, other factors such as the risk of predation, the rate of food intake  
374 (Pyke, 1984) or the densities of previous and simultaneous foragers (Lázaro et al., 2011) could  
375 also be influential, and therefore they should be examined to gain a comprehensive  
376 understanding of simultaneous bumblebees patch occupancy. Indeed, based on our field  
377 observations, whereas the other two plant species were almost exclusively visited by  
378 bumblebees during the recording periods, not only *B. terrestris* visited the *Persicaria* patches  
379 but other large-bodied insect taxa, such as flies and other bees, most likely changing  
380 bumblebees' attraction to flowers. Furthermore, the attractiveness of flower heads for foraging  
381 bumblebees changes rapidly within a patch, for instance with the amount of available nectar

382 and pollen (Somme et al., 2015) or the speed of rewards replenishment. Additionally, in the  
383 case of *Trifolium*, the cultivar can also influence the amount of nectar produced, thereby  
384 affecting its attractiveness to visiting insects (Szabo & Najda, 1985). This many sources of  
385 variability may explain why we did not detect significant differences between flower species  
386 when random variables were included and why results were different when simple regression  
387 was used. Thus, although it would be important to determine the influence of plant species on  
388 bumblebee carrying capacity, the high dynamism of the system makes this task extremely  
389 difficult if not impossible.

#### 390 4.3. The time spent with handling

391 Bumblebees spend a longer proportion of their total time on flowers ('bumblebee-time') relative  
392 to non-flower areas on *Trifolium*, compared to *Lotus* and *Persicaria*, likely due to differences  
393 in flower head size, resulting in longer handling time on plants with larger heads than on those  
394 with smaller ones. However, other factors like flower-specific parameters (e.g. structural  
395 complexity (Harder, 1983), nectar concentration and flower depth (Harder, 1986), and the  
396 nectar secretion rates (Stout & Goulson, 2002) can also influence how bumblebees optimise  
397 behaviour and thus handling time.

398 The probability of detecting a bumblebee on flower heads was higher on *Trifolium* compared  
399 to *Lotus* and *Persicaria*. This was likely because *Trifolium* flower heads are large, requiring a  
400 longer handling time, and thus, an increased probability of detecting a bumblebee on flowers.  
401 Moreover, the differences in time spent on flowers can also be the result of different strategies  
402 bumblebees choose to move between flowers. Since *Lotus* covers the surface in almost two  
403 dimensions, bumblebees tend to crawl between flowers (walk from one flower head to the next)  
404 but they tend to fly between *Trifolium* flowers, where flower heads are more scattered and vary  
405 in flower heights. Indeed, crawling between flowers is preferred as it expends significantly less  
406 energy compared to flight (Krell, 2018). Thus, since flying over non-flowery *Trifolium* areas is  
407 faster than crawling between flowers in the patches of the other two plants, the shorter off-

408 flower time of bumblebees on *Trifolium* can, at least partially, be explained. Our field  
409 observations, especially in the case of *Lotus* with a higher flower cover, supported this theory.  
410 Yet, bumblebees may need to make multiple flights between flower heads on the *Trifolium*  
411 patch (which is faster), whereas crawling between flower heads results in slower but  
412 uninterrupted means of transport on *Lotus*. This suggests that whilst the modes of movement  
413 differ, the overall distance covered by foraging individuals between flower heads can be similar.  
414 In summary, based on our results, we cannot conclusively support our hypothesis that there are  
415 significant differences in the time bumblebees spent on flowers (handling time) among the three  
416 plant species. Further research is needed to fully explore the foraging practices of bumblebees  
417 and understand how they optimize the time spent handling flowers.

#### 418 [4.4. Methodological perspectives](#)

419 In this study, we also tested the efficiency of a video-based recording method combined with  
420 computer vision analysis for studying bumblebee behaviour. Although our results are quite  
421 promising and we were able to detect bumblebees under field conditions on all three plant  
422 species with high accuracy, the field setting came with several challenges. Shadows, moving  
423 backgrounds, and varying light conditions can affect efficient object detection, and lead to false  
424 positive detections (Ratnayake et al., 2021). These, however, can be mitigated by improving  
425 training datasets, applying post-processing bounding box filters, and background subtraction  
426 techniques (Bjerge et al., 2024). Moreover, in natural vegetation, the 3D structure of plants (e.g.  
427 varying heights) can cause substantial difficulties.

428 Furthermore, detecting small insects in high-resolution images remains a challenge for current  
429 object detection models, but current advancements (e.g. the latest iteration in the YOLO series,  
430 Yolov8 (Jocher et al., 2023) with Slicing Aided Hyper Inference (SAHI) (Akyon et al., 2022))  
431 are enhancing the efficacy of these methods and making image pre-processing (such as tiling)  
432 unnecessary. Indeed, in our case, adjusting the camera height to ensure that the quadrat fits  
433 within the recorded area while keeping insects recognizable to the model was essential.

434 Additionally, physical quadrats disturbed foraging bumblebees and interfered with the image  
435 analysis (particularly during colour filtering), therefore, at later stages we used digitally  
436 designated quadrats, calibrated in post-processing, to avoid these issues. Despite these  
437 difficulties, we demonstrated promising results and we are confident that by individually  
438 tracking animals, these computer vision-based methods can be effective tools in providing new  
439 insight into previously unexplored or controversial behavioural issues of bumblebees.

#### 440 4.5. Future perspectives

441 The foundational principles of studying bumblebees and other pollinators were established as  
442 early as the middle and late 1990s, for example with a boom in research on 'optimal foraging  
443 theory' (Pyke, 1984). With the recent advancements of AI and computer vision, we now have  
444 the opportunity to revisit and improve these foundational studies with greater precision and  
445 larger sample sizes without exponentially increasing human labour. These novel technologies  
446 can facilitate more comprehensive and accurate studies of behavioural ecology, supporting  
447 biodiversity conservation and addressing many of the unanswered questions from earlier  
448 research. Nevertheless, further improvements are necessary to enhance the accuracy of insect  
449 detection and reliably capture fine-scale changes in behaviour. Studies like ours provide  
450 foundational insights that can inform future research without concerns about animal welfare  
451 (Lövei & Ferrante, 2024) or promoting cost-effective agricultural practices aligned with  
452 sustainability and conservation agriculture (e.g. in selecting plant species for landscape design).

453

#### 454 **Author Contributions**

455 Conceptualization, Z.V.S., G.S. and G.P.; Methodology, Z.V.S. and G.P.; Software, Z.V.S. and  
456 G.P.; Validation, Z.V.S. and G.P.; Formal Analysis, Z.V.S. and G.P.; Investigation, Z.V.S. and  
457 G.P.; Resources, Z.V.S. and G.P.; Data Curation, Z.V.S. and G.P.; Writing – Original Draft  
458 Preparation, Z.V.S. and G.P.; Writing – Review & Editing, Z.V.S., G.S. and G.P.;  
459 Visualization, Z.V.S. and G.P.; Supervision, G.P. and G.S.; Project Administration, Z.V.S.;

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461 the manuscript.

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## 467 **Data Availability**

468 The underlying computer code is available in the GitHub repository  
469 [https://github.com/zsvargaszilay/exploring\\_foraging\\_behaviour\\_with\\_computer\\_vision](https://github.com/zsvargaszilay/exploring_foraging_behaviour_with_computer_vision)

## 470 **Conflicts of Interest**

471 The authors declare no conflict of interest.

472

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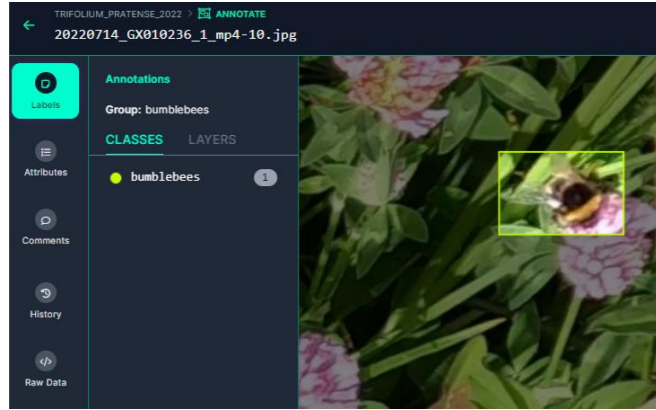
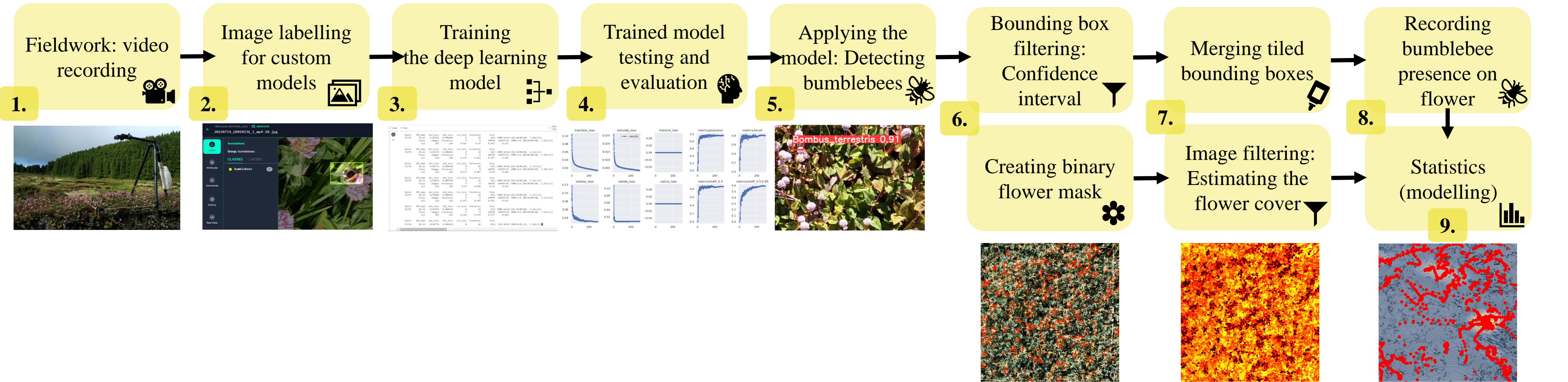
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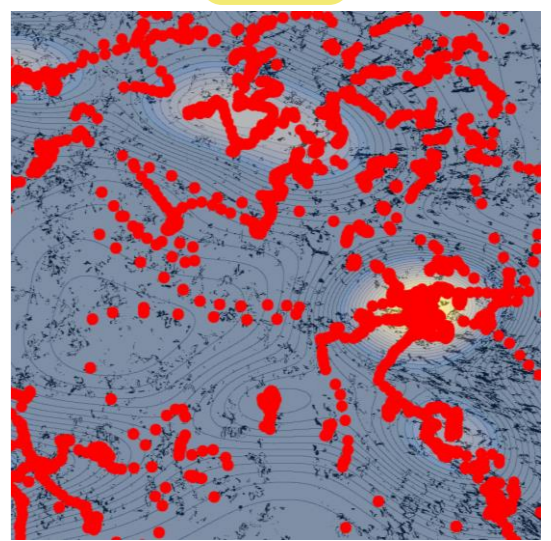
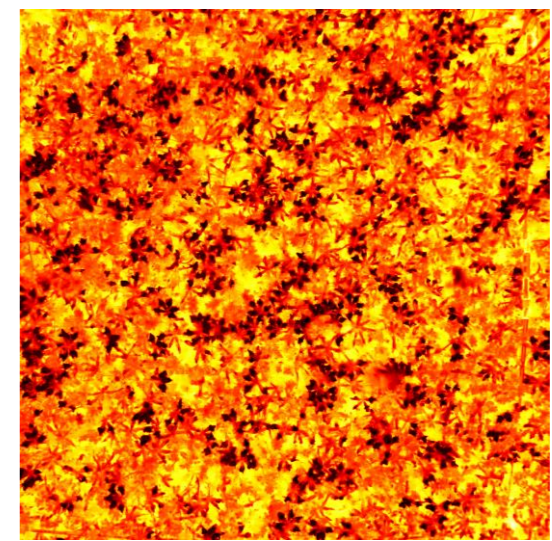
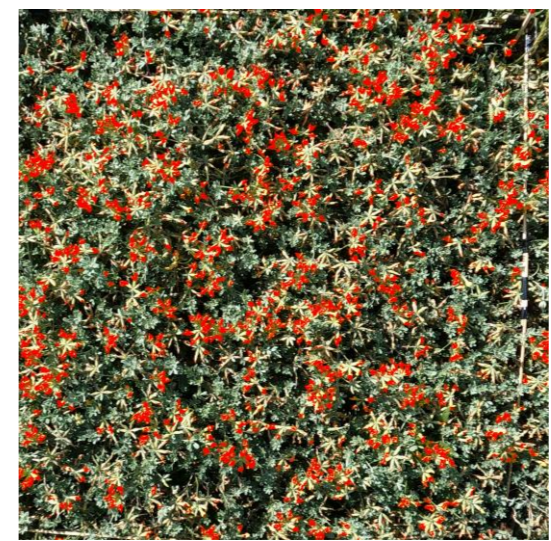
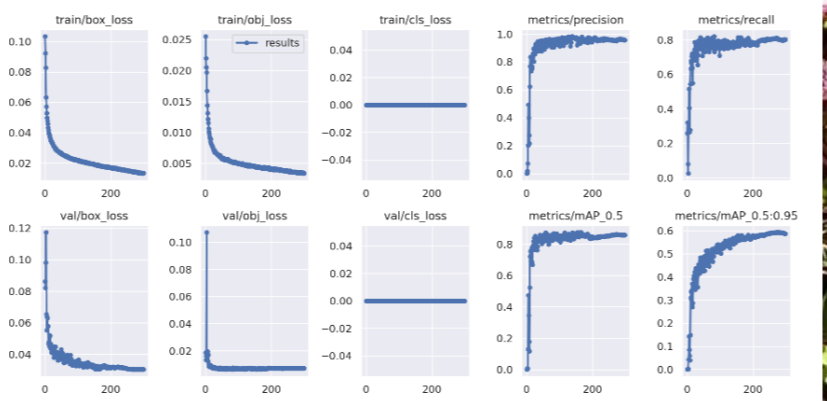
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Class  Images  Instances  AP@50  AP@75  AP@95
all     226     226     0.967  0.92  0.963  0.871
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Epoch  GPU_mem  box_loss  obj_loss  cls_loss  Instances  Size
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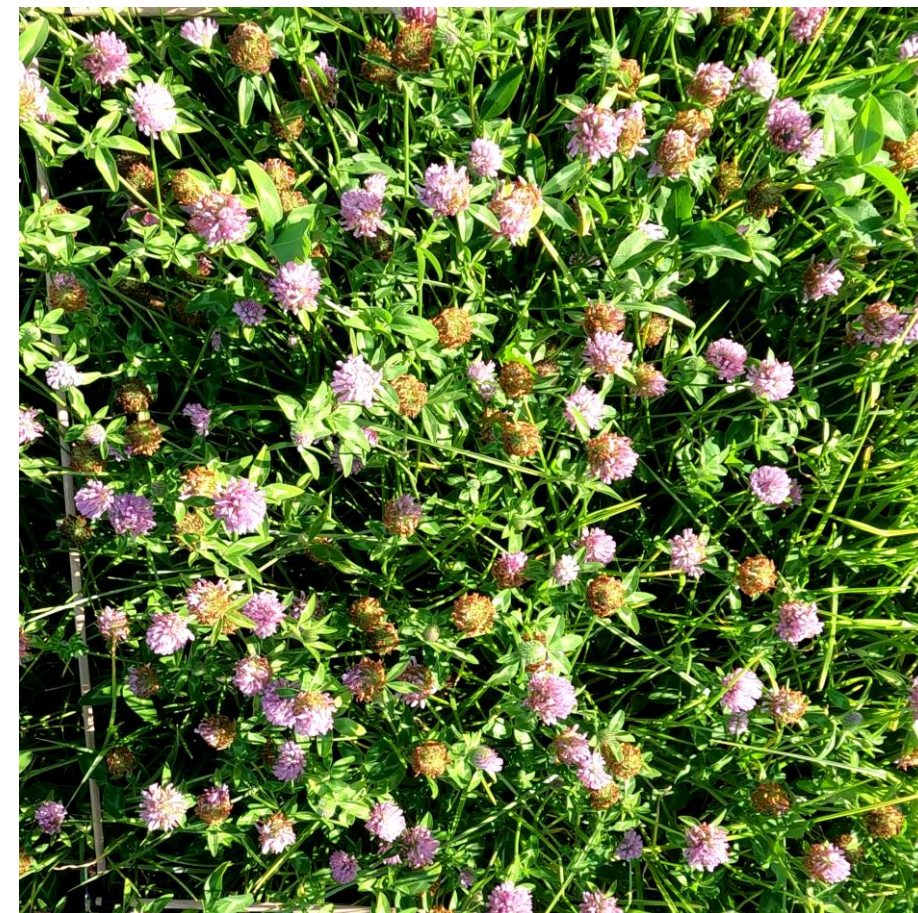
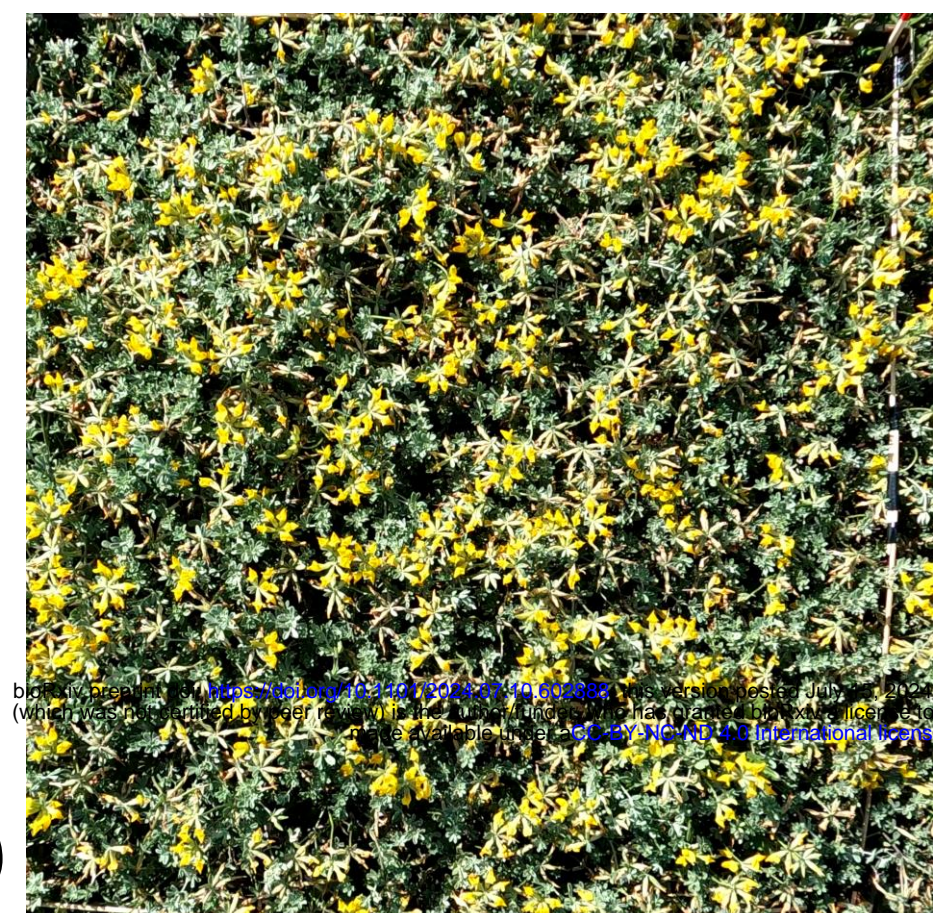
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*Lotus creticus*

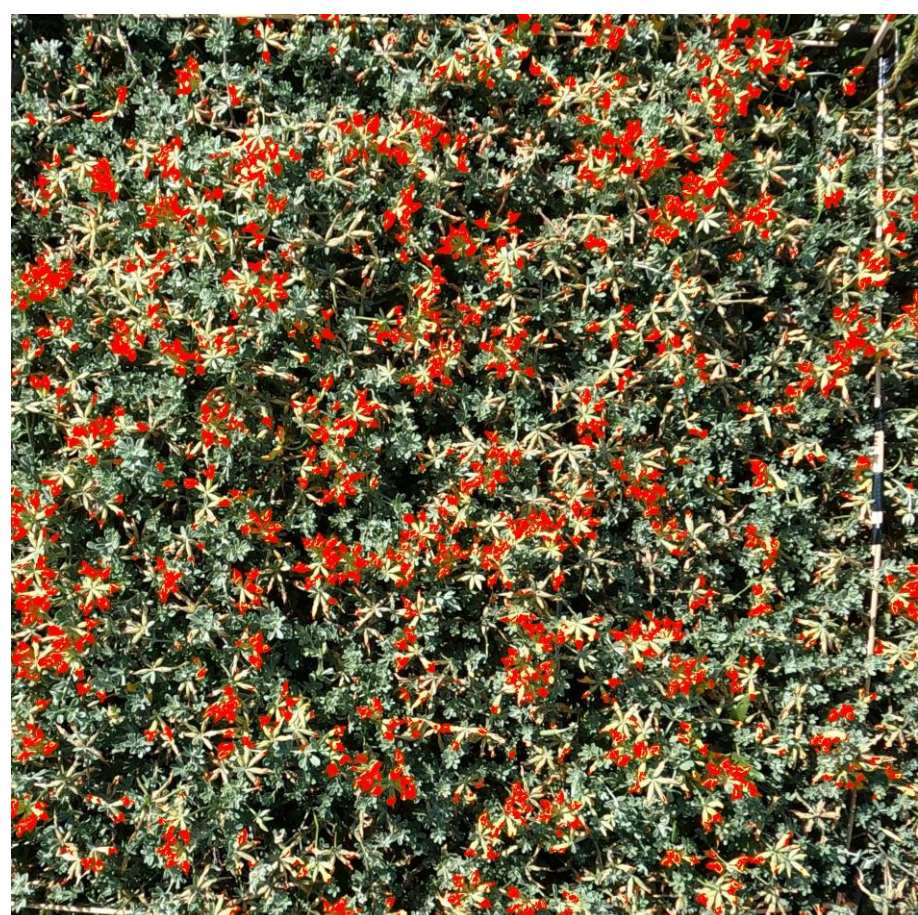
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*Trifolium pratense*

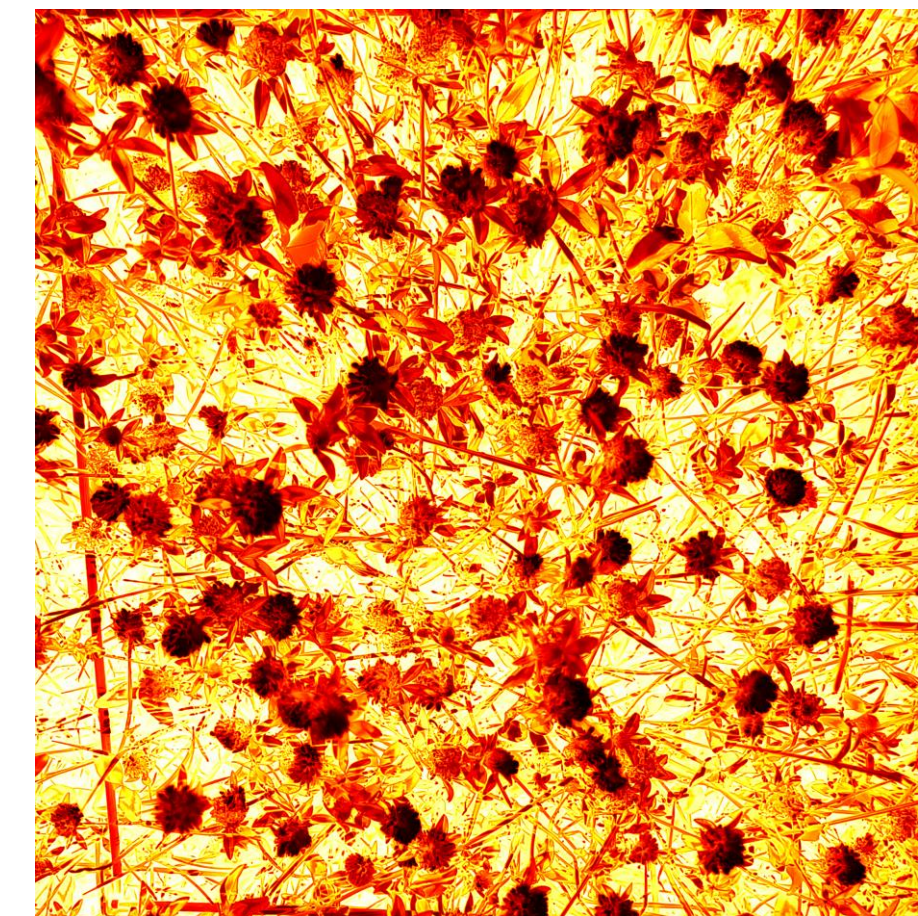
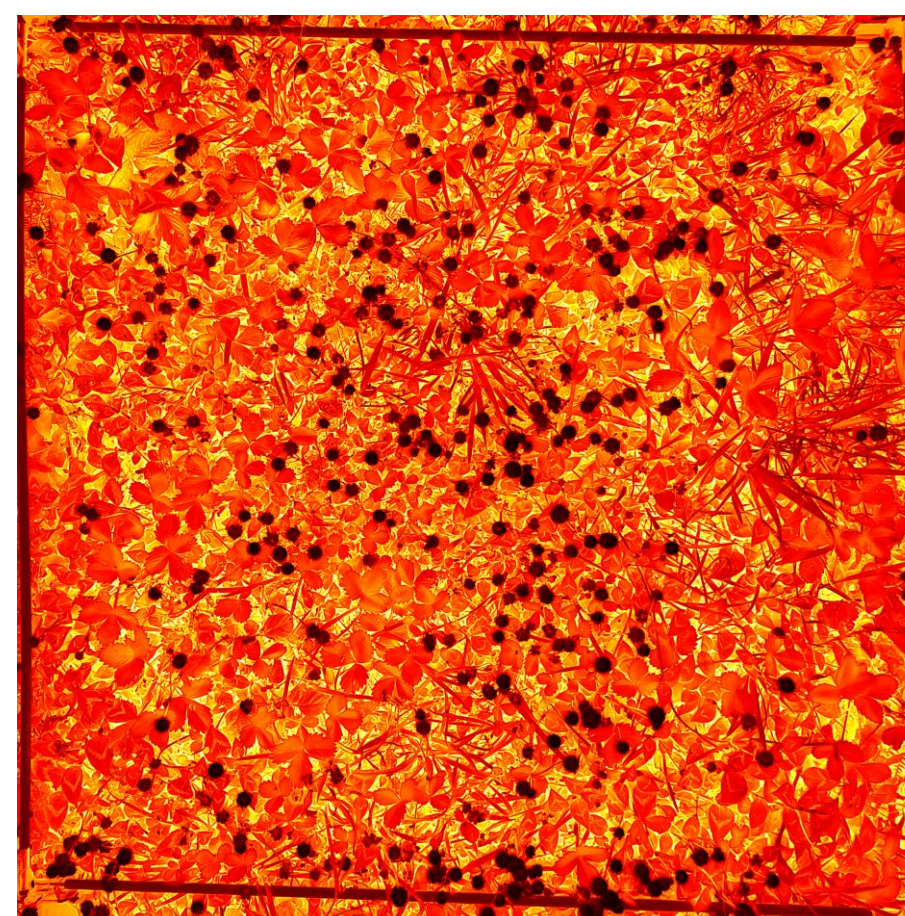
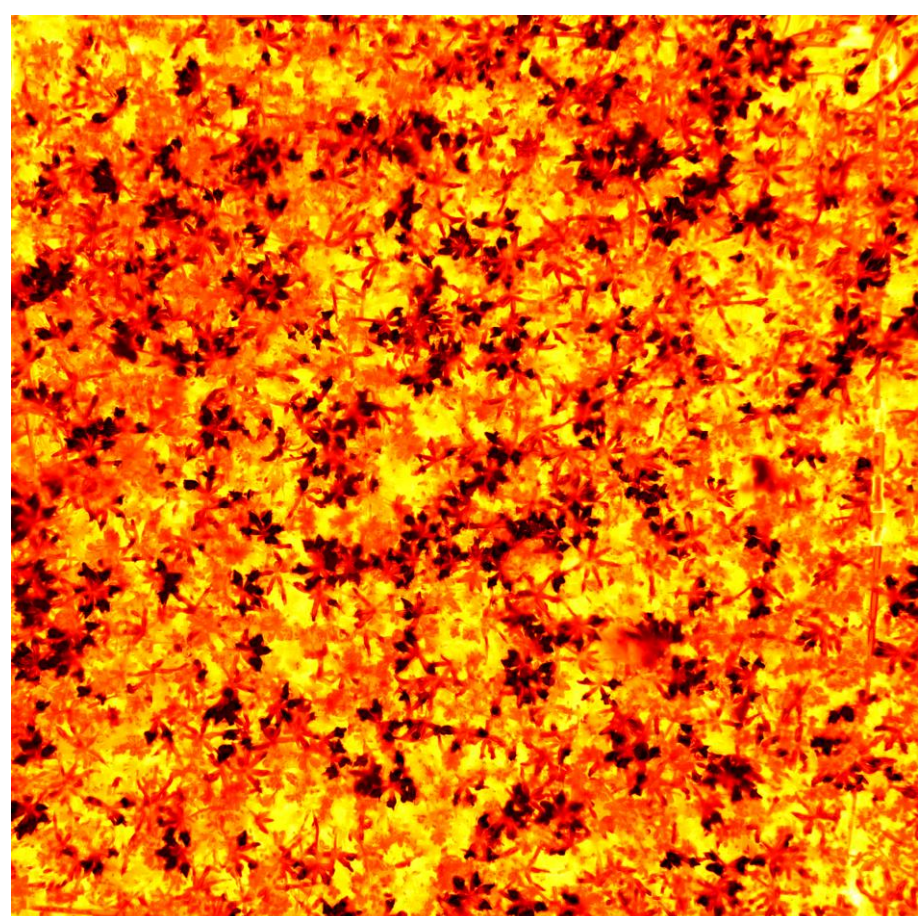


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(a)



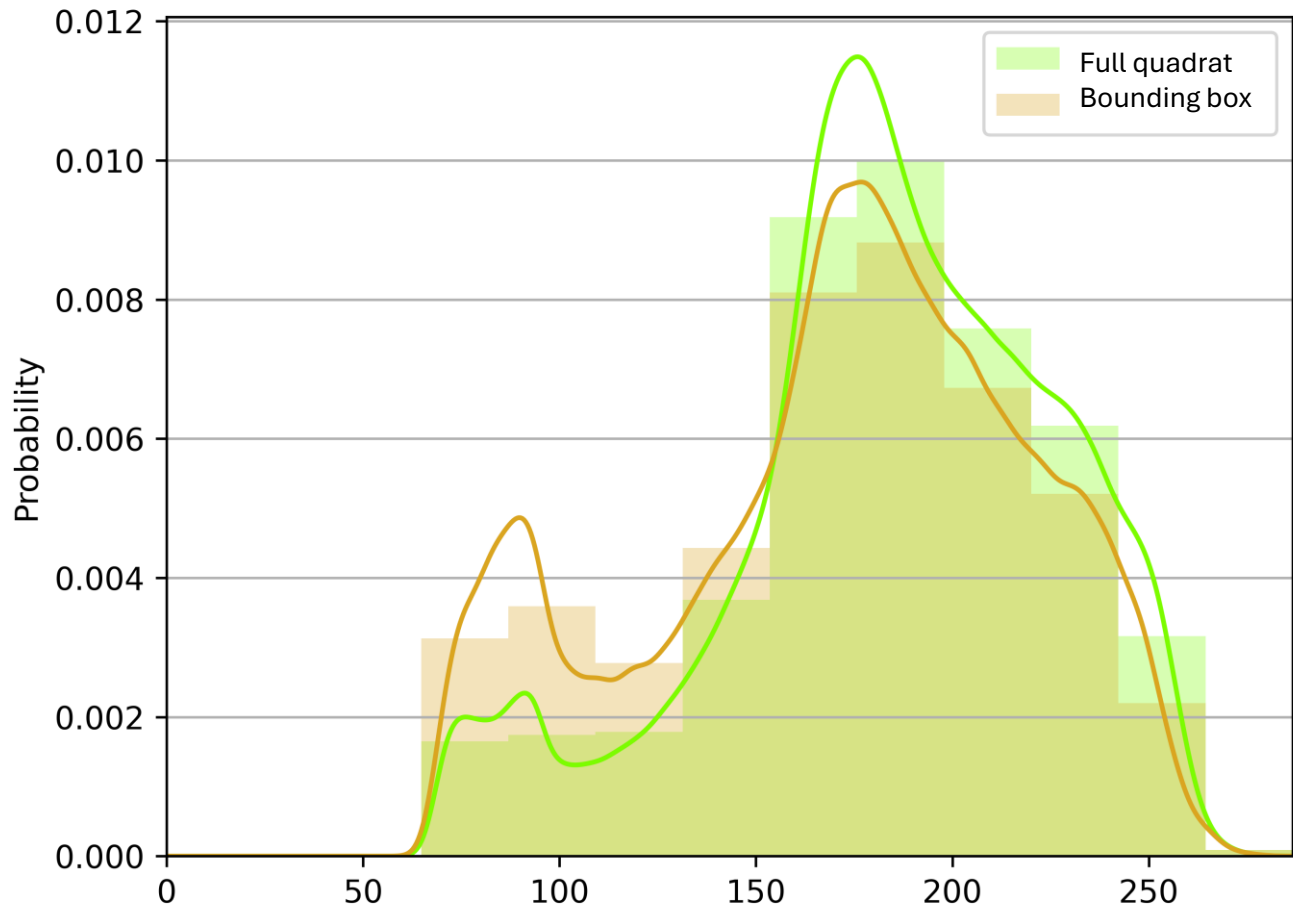
(b)



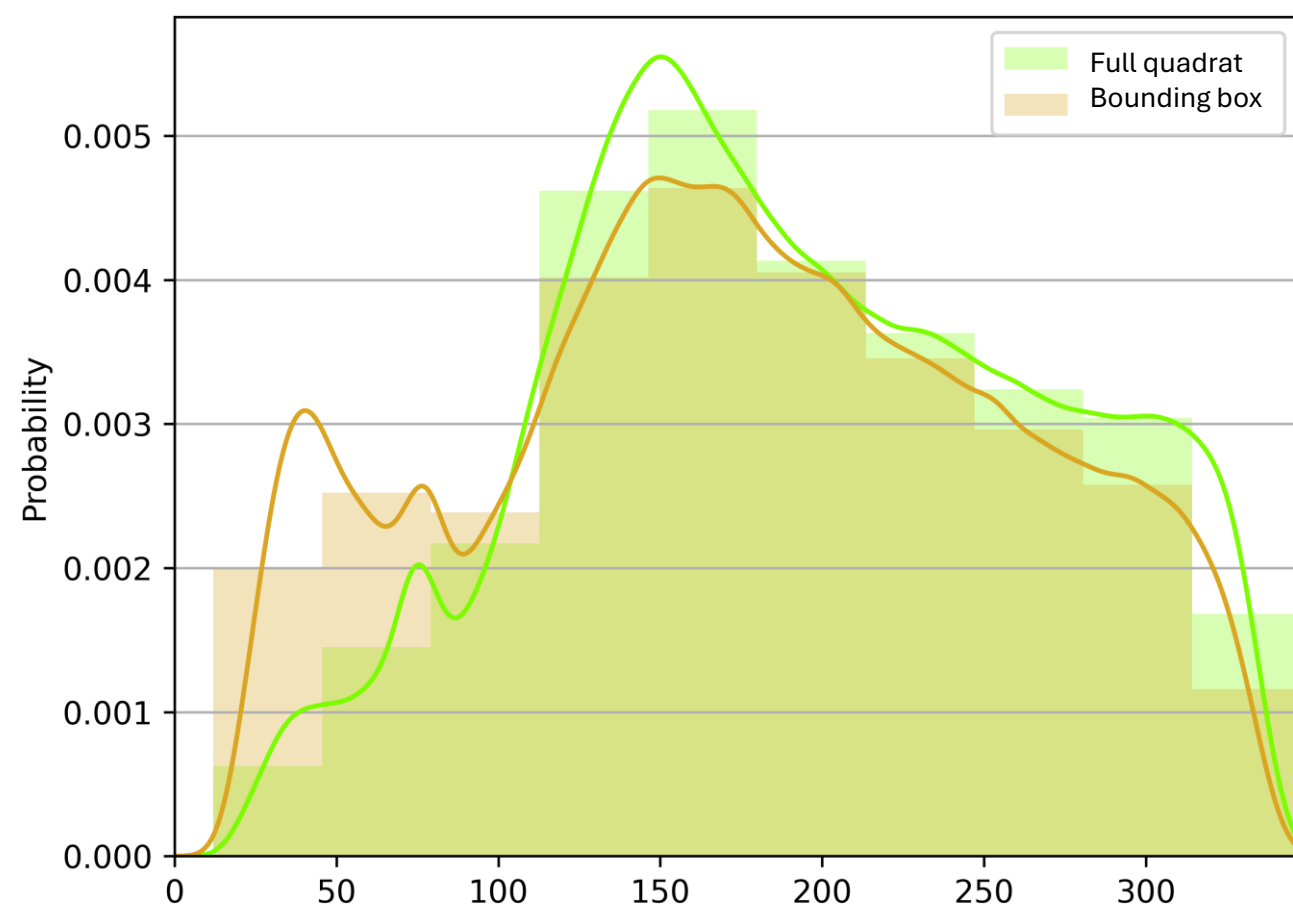
(c)

**(a)**

Lotus

**(b)**

Persicaria

**(c)**

Trifolium

